



Asymmetric synthesis of (2*S*,3*R*)- and (2*S*,3*S*)-[2-¹³C;3-²H] glutamic acid

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ABSTRACT

We have developed a synthetic route for (2*S*,3*R*)- and (2*S*,3*S*)-[2-¹³C;3-²H] glutamic acids with high enantioselectivity. The key reactions in this synthesis are the asymmetric reduction of the 2,3-didehydroornithine derivative using the (*S,S*)-Et-DuPHOS-Rh catalyst and the oxidation of the δ -position by ruthenium catalysis.

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The replacement of ¹H by ²H has been recognized as an efficient method for eliminating undesired peaks in ¹H nuclear magnetic resonance (NMR) spectra and for observing NMR signals against the relaxation caused by ¹H.¹ We recently proposed a novel labeling strategy, termed stereo-array isotope labeling (SAIL), for high-throughput NMR structure determination of proteins.² In the case of amino acid metabolism studies, the analysis of ¹³C isotope shifts induced by H/²H exchange in a ²H₂O system is utilized.³ In this study, we carried out the chemical synthesis of (2*S*,3*S*)- and (2*S*,3*R*)-[2-¹³C;3-²H] glutamic acids for metabolic analysis. Several groups have reported the stereoselective synthesis of glutamic acid derivatives labeled with deuterium at prochiral protons.⁴ However, these synthetic routes were not suitable for regioselective ¹³C labeling coupled with deuterium labeling because the starting ¹³C-labeled materials were unavailable. We therefore wish to report the simple synthesis of (2*S*,3*S*)- and (2*S*,3*R*)-[2-¹³C;3-²H] glutamic acids via the asymmetric hydrogenation of a 2,3-didehydroamino acid derivative, prepared from commercially available materials. Generally, during the synthesis of glutamic acid labeled with deuterium at the β -position, the synthetic route involving the hydrogenation of the 2,3-didehydroglutamic acid derivative may be straightforward. To the best of our knowledge, the regioselective ¹³C labeling of the 2,3-didehydroglutamic acid derivative can be achieved by using an α -oxoglutarate or a dehydroserine derivative as the starting material. This report describes a simple method for the synthesis of [2-¹³C;3-²H] glutamic acid, routed through ornithine as the key intermediate. Asymmetric hydrogenation of the [2-¹³C;3-²H] 2,3-didehydroornithine derivative,⁵ which is derived from readily available [2-¹³C] glycine and β -alanine, is carried out to yield an ornithine derivative labeled with deuterium at the β -position. This is stereoselectively followed by conversion of the

derivative into the target [2-¹³C;3-²H] glutamic acids by oxidation of the δ -position.

As shown in Scheme 1, the synthesis of (2*S*,3*R*)-[2-¹³C;3-²H] glutamic acid (**7**) commenced with the preparation of the stable isotope-labeled dehydroornithine derivative (**3**) using the Horner–Wadsworth–Emmons⁶ reaction. The starting *N*-Boc-3-amino[1-²H]propionaldehyde (**1**) was prepared by the reduction of the Weinreb amide⁷ of *N*-Boc- β -alanine with lithium aluminum deuteride (LiAlD₄),⁸ and was then condensed with phosphoryl[2-¹³C]glycine ester (**2**)⁹ in the presence of 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU) to give 2,3-didehydro[2-¹³C;3-²H]-ornithine (**3**) in 71% yield. Highly *Z*-selective olefination was observed when the reaction was conducted at –25 °C.

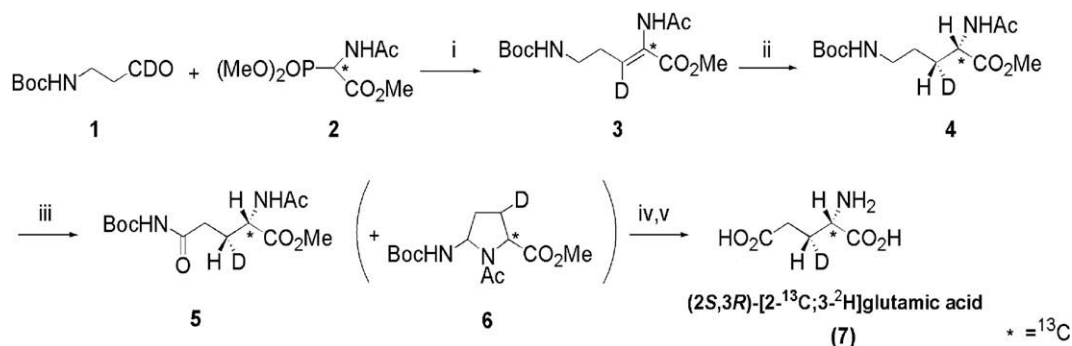
Asymmetric hydrogenation of the obtained dehydroornithine (**3**) was carried out under medium pressure (0.4 MPa) of hydrogen in the presence of (+)-1,2-bis[(2*S*,5*S*)-2,5-diethylphospholano]benzene (cyclooctadiene)rhodium(I) trifluoromethanesulfonate [(*S,S*)-Et-DuPHOS-Rh], because the DuPHOS family of catalysts is highly efficient for the hydrogenation of β -monosubstituted acetamidoacrylates, yielding a variety of amino acids with high enantioselectivities.¹⁰ In our previous study, a DuPHOS-Rh catalyst was also employed for the asymmetric hydrogenation of dehydroserine derivatives.¹¹ The stereochemistry at the α - and β -positions of [2-¹³C;3-²H]ornithine (**4**) was determined after the final glutamic acid was obtained.

According to Yoshifuji's procedure,¹² the conversion of ornithine (**4**) to glutamine (**5**) was performed using a catalytic amount of ruthenium dioxide and an excess of sodium periodate in a two-phase system of ethyl acetate and water. However, [2-¹³C;3-²H]glutamine (**5**) was obtained in only 38% yield because of the considerable side reaction yielding the cyclic compound **6**.

Hydrolysis of **5** was performed by refluxing it in 2 M HCl, and subsequent ion exchange treatment of the obtained glutamic acid hydrochloride with DOWEX 50WX8 afforded (2*S*,3*R*)-[2-¹³C;3-²H]

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Scheme 1. Reagents and conditions: (i) DBU, CH₂Cl₂, -25 °C, 71%; (ii) (*S,S*)-Et-DuPHOS-Rh, benzene, H₂ (0.4 MPa), 84%; (iii) RuO₂, NaIO₄, ethyl acetate-H₂O, 38%; (iv) 2 M HCl reflux; (v) DOWEX 50WX8, 70% (2 steps).

glutamic acid (**7**) in 70% yield. The enantiopurity based on the α -position was determined to be 99% ee by high performance liquid chromatography (HPLC) analysis using a chiral column (MCIGEL CRS10W). Figure 1a shows the 300 MHz ¹H NMR spectrum of **7**. As compared to that of the unlabeled glutamic acid (Fig. 1d), the signal for the 3R proton disappears completely, indicating that **7** has *R* stereochemistry at the β -position. This stereochemical outcome can be attributed to the exclusive *cis*-addition of hydrogen to the α -re-face of deuterated (*Z*)-2,3-didehydroornithine (**3**), promoted by the (*S,S*)-Et-DuPHOS-Rh catalyst.

We next performed the asymmetric deuteration of 2,3-didehydro[2-¹³C]ornithine (**9**), prepared from *N*-Boc-3-aminopropionaldehyde (**8**) and **2**, in 60% yield (Scheme 2), in order to obtain its isotopic stereoisomer, (2*S*,3*S*)-[2-¹³C;3-²H] glutamic acid (**13**).

The (*S,S*)-Et-DuPHOS-Rh catalyst afforded [2-¹³C;2,3-²H₂]ornithine (**10**) in 98% yield with the (2*S*,3*S*)-configuration, which was confirmed after its conversion to glutamic acid.

The obtained ornithine (**10**) was then subjected to ruthenium-catalyzed oxidation. Sheldon and co-workers pointed out that low pH has a detrimental effect on a catalytic system involving ruthenium tetroxide.¹³ We found that keeping the pH mildly alkaline by adding aqueous sodium carbonate during the course of the reaction improved the chemical yield of the glutamine from 38% (for **5**) to 64% (for **11**).

The glutamine (**11**) was then deprotected to give (2*S*,3*S*)-[2-¹³C;2,3-²H₂] glutamic acid (**12**) in 82% yield. The enantiopurity of **12** with respect to the α -position was also confirmed by HPLC analysis to be 99% ee. Its ¹H NMR spectrum (Fig. 1b) confirmed that

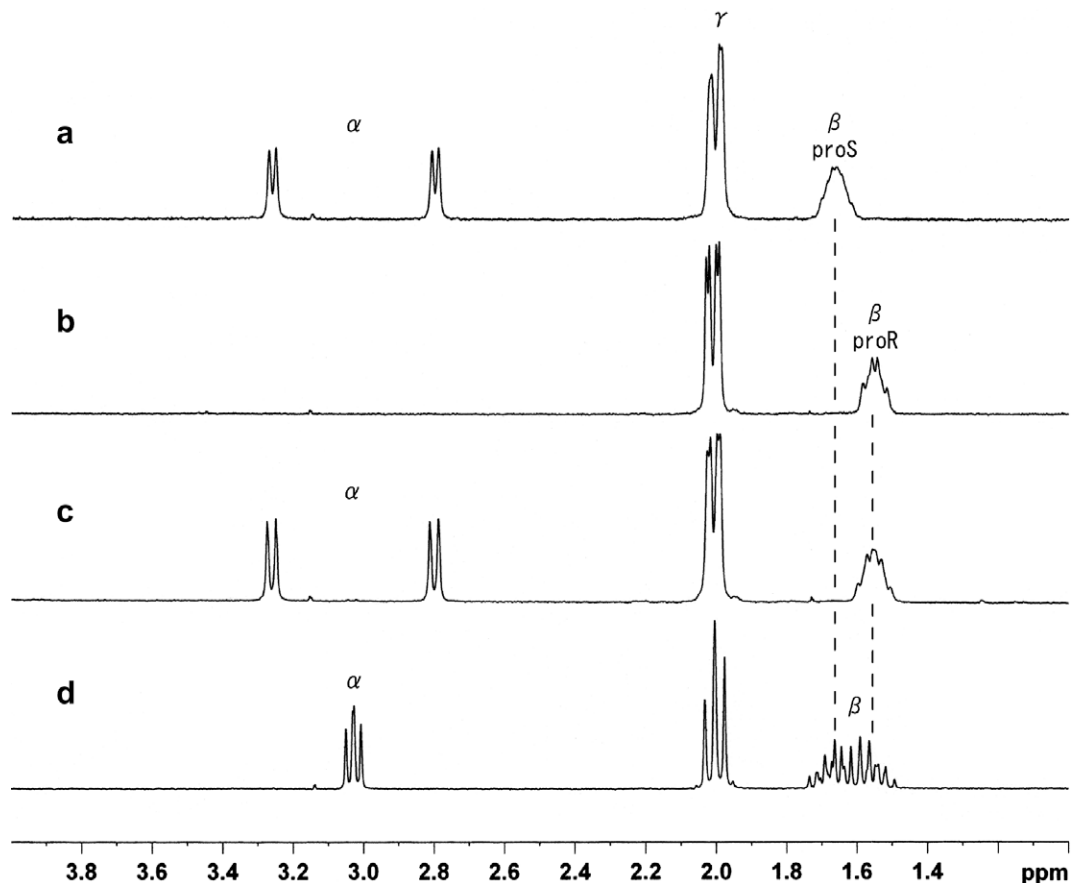
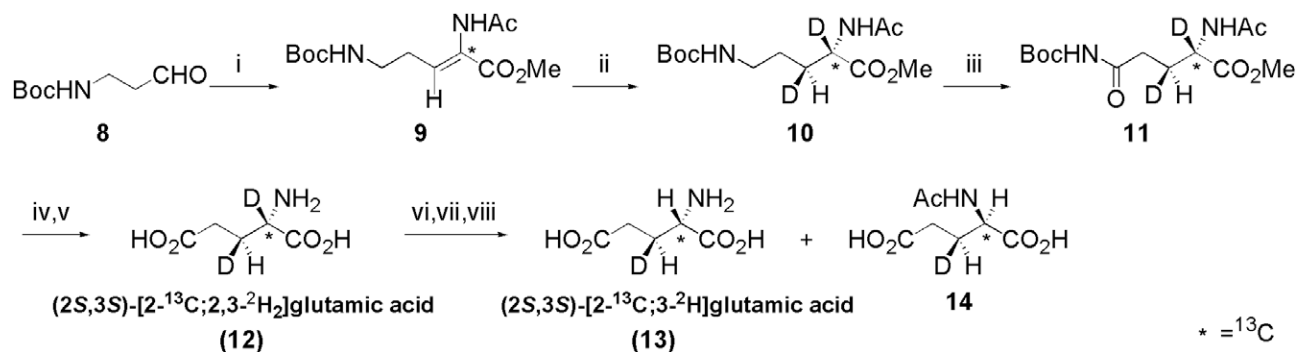


Figure 1. 300 MHz ¹H NMR spectra of (a) (2*S*,3*R*)-[2-¹³C;3-²H]glutamic acid (**7**), (b) (2*S*,3*S*)-[2-¹³C;2,3-²H₂]glutamic acid (**12**), (c) (2*S*,3*S*)-[2-¹³C;3-²H]glutamic acid (**13**), and (d) unlabeled glutamic acid in NaOD-D₂O.



Scheme 2. Reagents and conditions: (i) **2**, DBU, CH₂Cl₂, -25 °C, 83%; (ii) (*S,S*)-Et-DuPHOS-Rh, benzene, D₂ (0.4 MPa), 98%; (iii) RuO₂, NaIO₄, Na₂CO₃, ethyl acetate–H₂O, 64%; (iv) 2 M HCl reflux; (v) DOWEX 50WX8, 82% (2 steps); (vi) 5 M NaOH, (CH₃CO)₂O, 38 °C; (vii) porcine kidney acylase I, pH 8, 37 °C; (viii) DOWEX 50WX8, (**13**): 39% (2 steps), (**14**):43%.

the regioselective and stereoselective incorporation of stable isotopes were accomplished. Finally, we carried out deuterium-hydrogen exchange at the α -position using a traditional racemization-resolution procedure.¹⁴ Thus, **12** was treated with an excess of acetic anhydride in 5 M NaOH at 38 °C overnight. The optical resolution of the racemic *N*-acetylglutamic acid was effected through the L-directed deacetylation with porcine kidney acylase I to give (2*S*,3*S*)-[2-¹³C;3-²H] glutamic acid (**13**) in 39% yield, along with unreacted (2*R*,3*S*)-*N*-acetyl[2-¹³C;3-²H] glutamic acid (**14**) in 43% yield. The ¹H NMR spectrum of **13** (Fig. 1c) indicated the complete replacement of α -deuterium by a hydrogen atom.

In summary, we have achieved the asymmetric synthesis of (2*S*,3*R*)- and (2*S*,3*S*)-[2-¹³C;3-²H] glutamic acids with high enantioselectivity. The key reactions in this synthesis are the asymmetric hydrogenation or deuteration of the 2,3-didehydroornithine derivative using the (*S,S*)-Et-DuPHOS-Rh catalyst and the subsequent ruthenium-catalyzed δ -oxidation leading to the formation of the glutamine derivative. Further modification of the present procedure to obtain other labeled amino acids is now underway.

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References and notes

- Butter, W. T. *Science* **1968**, *161*, 795–798; Markley, J. L.; Putter, I.; Jardetzky, O. *Science* **1968**, *161*, 1249–1251.
- Kainosho, M.; Torizawa, T.; Iwashita, Y.; Terauchi, T.; Ono, A.; Güntert, P. *Nature* **2006**, *440*, 52–57.
- García-Martín, M. L.; García-Espinosa, M. A.; Ballesteros, P.; Bruix, M.; Cerdán, S. *J. Biol. Chem.* **2002**, *277*, 7799–7807.
- Dietrich, P.; Young, D. W. *Tetrahedron Lett.* **1993**, *34*, 5455–5458; (b) Oba, M.; Terauchi, T.; Hashimoto, J.; Tanaka, T.; Nishiyama, K. *Tetrahedron Lett.* **1997**, *38*, 5515–5518.
- Baldwin, J. E.; Merritt, K. D.; Schofield, C. J. *Tetrahedron Lett.* **1993**, *34*, 3919–3920.
- Boutagy, J.; Thomas, R. *Chem. Rev.* **1974**, *74*, 87–99.
- Nahm, S.; Weinreb, S. M. *Tetrahedron Lett.* **1981**, *22*, 3815–3818.
- Blaney, P.; Grigg, R.; Rankovic, Z.; Thornton-Pett, M.; Xu, J. *Tetrahedron* **2002**, *58*, 1719–1737.
- (a) Mazurkiewicz, R.; Kuźnik, A.; Grymel, M.; Październiak-Holewa, A. *ARKIVOK* **2007**, *vi*, 193–216; (b) Schmidt, U.; Lieberknecht, A.; Wild, J. *Synthesis* **1984**, 53–60; (c) Oba, M.; Ueno, R.; Fukuoka, M.; Kainosho, M.; Nishiyama, K. *J. Chem. Soc., Perkin Trans. 1* **1995**, 1603–1609.
- (a) Burk, M. J. *Acc. Chem. Res.* **2000**, *33*, 363–372; (b) Burk, M. J.; Feaster, J. E.; Nugent, W. A.; Harlow, R. L. *J. Am. Chem. Soc.* **1993**, *115*, 10125–10138; (c) Stammers, T. A.; Burk, M. J. *Tetrahedron Lett.* **1999**, *40*, 3325–3328; (d) Jones, S. W.; Palmer, C. F.; Paul, J. M.; Tiffin, P. D. *Tetrahedron Lett.* **1999**, *40*, 1211–1214; (e) Debenham, S. D.; Debenham, J. S.; Burk, M. J.; Toone, E. J. *J. Am. Chem. Soc.* **1997**, *119*, 9897–9898; (f) Rizen, A.; Basu, B.; Chattopadhyay, S. K.; Dossa, F.; Frejd, T. *Tetrahedron: Asymmetry* **1998**, *9*, 503–512; (g) Hiebl, J.; Kollmann, H.; Rovenszky, F.; Winkler, K. *J. Org. Chem.* **1999**, *64*, 1947–1952.
- Terauchi, T.; Kobayashi, K.; Okuma, K.; Oba, M.; Nishiyama, K.; Kainosho, M. *Org. Lett.* **2008**, *10*, 2785–2787.
- Yoshifuji, S.; Tanaka, K.; Nitta, Y. *Chem. Pharm. Bull.* **1987**, *35*, 2994–3001.
- Gonsalvi, L.; Arends, I. W. C. E.; Sheldon, R. A. *Chem. Commun.* **2002**, 202–203.
- Greenstein, J. P.; Winitz, M. *Chemistry of the Amino Acids*; John Wiley and Sons: New York, 1961.